Reactions with and in anhydrous hydrogen fluoride systems. Part 8^* . Triethylamine trishydrofluoride - a convenient reagent for the stereoselective synthesis of glycosyl fluorides

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Abstract

Stereoselective fluorinations are effected by bromine-fluorine exchange at glycosyl bronudes of the D- and L-series using triethylamine trishydrofluoride. F'yranosyl fluorides of D-Xylose, L-arabinose, D-glucose, D-mannose and L-rhamnose derivatives, as well as of D**galacturonic acid esters, have been prepared. The influence of neighbouring groups in the 2-position is considered.**

Introduction

Glycosyl fluorides may be of interest for glycosylations [2-41. Anomerically pure glycosyl fluorides are of greatest importance, but many fluorination methods lead to anomeric mixtures [2]. Good selectivities are obtained by bromine-fluorine exchange at protected glycosyl bromides with the help of various fluorinating reagents [2, 51. Recently, we found that the two-phase system $Et_3N \cdot 3HF/CCl_4$ is a convenient reagent for selective bromine-fluorine exchange with inversion at the anomeric centre of sugar derivatives [6a]. The advantages of the reagent result from a smaller expenditure of time and/or cost compared with other reported methods. Some additional investigations have now been carried out with this reagent to demonstrate the influence of neighbouring groups in the 2-position of a sugar molecule on the selectivity of fluorination.

^{*}For part 7, see ref. 1.

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Results and discussion

 $2,3,4$ -Tri-O-acetyl- α -D-xylopyranosyl- (1) [7a], $2,3,4$ -tri-O-acetyl- β -L-arabinopyranosyl- (3) [7b], 2,3,4,6-tetra-O-methyl- α -D-glucopyranosyl- (5) [7c], 2,3,4,6-tetra-O-benzoyl-a-D-mannopyranosyl- (7) [7d] and 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl bromide (9) [7e] as well as methyl- (11) [7f] and benzyl- $(2,3,4\text{-tri}-O\text{-}aeetyl-\alpha-D\text{-}galactopyranosyl bromide)uronate (13) [7g] were fluor$ inated under the same conditions as described in ref. 6a. Heating (reflux) of the pentosyl bromides 1, 3 $(0-5 h)$ and hexosyl bromides 5, 7, 9, 11, 13 (2 h) in the $Et_3N \cdot 3HF/|CCl_4$ system (1:5-6 v/v) gave the corresponding glycosyl fluorides (Table 1, Schemes 1 and 2). No anomerisation of the product glycosyl fluorides was observed with $Et_3N \cdot 3HF/|CCl_4$. On the other hand, the same conditions were unable to effect the conversion of 3-O-(nalkyl)-2,4,6-tri-O-acetyl- α -D-glucopyranosyl bromides with long chains (dodecyl or hexadecyl) into the corresponding β -fluorine derivatives [6c]. The high lipophilicity of such compounds and the shielding of the reaction centre by the alkyl chain hinder reagent attack.

With the exception of compounds 4 and 14, the glycosyl fluorides have been previously described in the literature, but their NMR data (Table 2) have only been partially published except for $2.3.4$ -tri-O-acetyl- β -D-xylopyranosyl fluoride (2) [12, 13]. The glycosyl bromides 11 and 13 are very unstable and were not isolated before fluorination (see example 2 in the Experimental section); the ester groups are also easily cleaved in 13 and 14. All the noted yields are related to the work-up of the Cl_4 phases and the pure products (crude products in parentheses, Table 1); the other phase contained by-products (partially deprotected sugars).

The NMR data for D -xylopyranosyl fluoride (2) are identical with the literature values [12, 13]. The ¹H spectrum of $2,3,4$ -tri-O-acetyl- α -L-arabinopyranosyl fluoride (4) was the same as that of the $2,3,4$ -tri-O-acetyl- α -Darabinopyranosyl fluoride $(^4C_1$ conformation) described in ref. 12. Because

TABLE 1

"See Schemes 1 and 2.

bEther/pentane.

 $C₂,3,4$ -Tri-O-acetyl- α -D-arabinopyranosyl fluoride (cf. 4) [8, 12].

Scheme 1.

the coupling constants $J_{H-4, H-5a}$ and $J_{H-4, H-5e}$ are also the same as in ref. 12, the acetyl group in the 4-position cannot be axial but must be equatorial in agreement with a ${}^{1}C_{4}$ conformation for 4. The relatively large coupling constants between the C-2 atom and fluorine in 2 and 4 $(J_{C_2,F} = 36$ and 34.8 Hz) provide a further reason for believing that the fluorine has an axial configuration. The 13C NMR spectra of compounds 12 and 14 confirm the p-structure of these fluorides through chemical shifts of 106.8 ppm for C-1 and by the coupling constants $J_{C-1, F} = 221$ Hz and $J_{C-3, F} = 10.7$ -10.8 Hz (Table 2). The literature value [5c] quoted for δ_{H-1} for compound 12 is not correct; we found that for both 12 and 14, $\delta_{H-1} = 5.28$ (dd, $J_{H-1, H-2} = 7.0-7.2$) Hz) ppm.

The experimental results, including those of ref. 6, lead to the following conclusions:

Competing attacks:

(A) trans attack of a fluoride ion on 9 producing 10β

(B) trans attack of the 2-O-acetyl group, following an attack of a fluoride ion producing 10α Scheme 2.

- 1. Bromine-fluorine exchange takes place selectively with inversion and formation of only one anomer if the leaving group and the group in the 2-position are *cis*-orientated; the attack of the fluoride ion can take place from the *trans*-direction corresponding to a real S_n 2-type reaction.
- 2. When a *trans* configuration exists for both groups, one obtains anomeric mixtures $(8, 10)$ because the fluoride ion competes in an S_N2 -type reaction with the lone pairs of the oxygen at the neighbouring group, which can form a cyclic oxonium ion (Scheme 2). The nucleophilicity of the fluoride ion in the corresponding fluorination system is decisive for the selectivity. Conformation effects support the formation of an α -anomer in the case of manno-configurated sugars $(\Delta 2 \text{ effect } |14|).$

Experimental

All the glycosyl bromides were prepared using the HBr/acetic acid procedure described in ref. 7g. The properties of the several bromides are described in refs. 7a-g; see marked reactants in Table 1. Fluorination was controlled by the use of TLC methods using Alufolie Kieselgel 60 F 254 (Merck) and the solvent system toluene/ethyl acetate $= 3:1$. Triethylamine trishydrofluoride was also a commercial product, but the procedure described below was used to prepare the complete reagent system *in situ.*

NMR spectra using CDCl₃ solutions were obtained on a Bruker WM 250 MHz instrument; see Table 2.

Triethylamine trishydrofuoride/carbon tetrachloride system

Dried triethylamine (13.8 ml, 0.1 mol) in CCl_4 (100 ml) was placed in a 250 ml glass bottle and cooled down to $\< 0$ °C. Anhydrous HF (6 ml, 0.3)

TABLE 2

¹H NMR and ¹³C NMR data (CDCl₃; TMS; δ in ppm; *J* in Hz)

mol) was then added dropwise directly to the solution using an HF-resistant dropping tube with stirring. The solution warmed up during the reaction and a second phase $(Et₃N. 3HF)$ formed. (Care! Do not contact glass with **anhydrous HF.)** The glycosyl bromides **1,** 3, 5, 7, 9, **11** or 13 (0.5-1.0 mmol) were then added directly or in CCI_4 to this reagent.

Example 1: (fluorination procedure)

 $2.3.4$ -Tri-O-acetyl- α / β -L-rhamnopyranosyl fluorides $10\alpha/10\beta$

2,3,4-Tri-O-acetyl- α -L-rhamnopyranosyl bromide (9) [7e] (1.77 g, 5 mmol) was added to Et₃N. 3HF (16 g, 0.1 mol)/CCl₄ (100 ml). After 2 h reflux^{*} with vigorous stirring, the phases were separated and extracted with 30 ml $CCl₄$. Finally, the combined $CCl₄$ phases were washed free from acid with water. After filtration and evaporation of the solvent the anomeric mixture of $10\alpha/\beta$ was obtained as a syrup. Yield after chromatographic purification, 0.6 g (41%); for analytical data see Tables 1 and 2.

Example 2: (combination of bromination /7gJ and fkwination procedures)

BtmzyL(2,3,4-tri-0-acetyl-a-13galactopyranosyl bromide)uronute (14)

Benzyl(1,2,3,4-tetra-O-acetyl- α/β -D-galactopyranose)uronate [15](3.17g, 7 mmol), dissolved in 10 ml CCL, was added to a 30% HBr solution in acetic acid (45 ml) at 0 °C. After 2 h standing at this temperature (TLC control), the reaction mixture was poured on ice (100 g) and extracted with CCl_4 (2-3 times). The combined CCl_4 phases were washed with ice water until neutral, dried and evaporated at 30-40 "C. The syrup obtained was dissolved in some CCL, and added directly as a crude product to the $Et₂N \cdot 3HF$ $(0.1 \text{ mol})/\text{CCl}_4$ (100 ml) system described above. With stirring and reflux, fluorination was complete after 2 h. Work-up was as described for **10 in** the fluorination procedure (Example 1). The syrup obtained was dissolved in ether and pentane was added until precipitation of crystalline 14 occurred. Yield, 1.15 g (40%); m.p., 167–169 °C (recrystallisation from ethyl acetate/ hexane); $\left[\alpha\right]_D^{22} = +28.8$ (c=0.9, CHCl₃); further analytical data in Tables 1 and 2.

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References

- 1 Part 7: R. Miethchen, T. Gabriel and G. Kolp, *Synthesis,* (1991) 885.
- 2 (a) F. Micheel and A. Klemer, *Adv. Ccwbohydr. Ch_em., I6* (1961) 85; (b) A. A. E. Penglis, *Adv. Carbohydr. Chem. Biochem., 38 (1981) 195; (c) R. Miethchen, Z. Chem., 29 (1989)* 425; (d) T. Tsuchiya, *Adv. Cm-bohydr. Chem. Biochem., 48* (1990) 91.
- 3 H. Waldmann, *Nachr. Chem. Tech. Lab., 39* (1991) *675.*
- *4* J. Jiinnemann, I. Lundt and J. Thiem, *Liebigs Ann. Cha.,* (1991) 759.

^{*}Pentose derivatives required 0.5 h reflux

- 5 (a) Ya. V. Voznyi, I. S. Kalichewa and A. A. Galoyan, Biomg. Chim., 7 (1981) 406; (b) L. D. Hall, J. F. Manville and N. S. Bhacca, Can. J. Chem., 47 (1969) 1; (c) M. Kreuzer and J. Thiem, Carbohydr. Res., 149 (1986) 347; (d) J. Jünnemann, I. Lundt and J. Thiem, *Acta Chem. Scan&, 45* (1991) 494.
- 6 (a) R. Miethchen, G. Kolp, D. Peters and J. Holz, Z. Chem., 30 (1990) 56; (b) G. Kolp, *Dissertation (thesis),* University of Restock, 1991; (c) J. Holz, *Dissertation (thesis),* University of Restock, 1992.
- 7 (a) R. L. Whistler and M. L. Wolfrom, *Methods in Carbohydrate Chemistry,* Academic Press, New York, London, 1962, Vol. I, p. 182; (b) R. E. Deriaz, W. G. Overend, M. Stacey, E. G. Teece and L. F. Wiggins, J. *Chem. Sot.,* (1949) 1879; (c) H. A. Bates and C. Selick, J. *Curbohydr. Chem., 4* (1985) 273; (d) R. K. Ness, H. G. Fletcher and C. S. Hudson, J. *Am. Chem. Sot., 72* (1950) *2200; (e)* E. Fischer, M. Bergmann and A. Rabe, *Ber. Dtsch. Chem. Ges., 53 (1920) 2362; (f) E. L. Pippen and R. M. McCready, J. Org. Chem., 16* (1951) 262; (g) V. I. Betaneli, M. V. Ovchinnikov, L. V. Backinowsky and N. K. Kochetkov, *Carbohydr. Res., 84* (1980) 211.
- 8 I. Lundt and C. Pedersen, *Microchim. Acta,* (1966) 126.
- 9 G. L. Trainor, J. *Curbohydr. Chem., 4* (1985) *545.*
- 10 C. Pedersen, *Acta Chem. Scund., 17* (1963) *673.*
- 11 R. W. Binkley, *Modern Carbohydrate Chemistry,* Marcel Dekker, New York, Basel, 1988, p. 53.
- 12 L. D. Hall and J. F. Manville, Can. J. Chem., 47 (1969) 19.
- 13 K. Bock and C. Pedersen, *Acta Chem. Scand., B29* (1975) 682.
- 14 J. Dale, *Stereochemie und Korzfmma tionsanalyse,* Verlag Chemie, Weinheim, New York, 1978, p. 155.
- 15 C. Vogel, H. Boye and H. Kristen, *J. Prakt. Chem.*, 332 (1990) 28.